

Cytopathological and Numerical Changes in Haemocytes of *Periplaneta americana* after Treatment with Malathion

S. C. Saxena and Prabhu N. Saxena*

Toxicology Laboratory, Department of Zoology, University of Rajasthan,
Jaipur - 302004, India

Haemolymph plays a significant role in the detoxification of chemicals. (Patton 1961) observed a reduction in haemocyte numbers after the application of parathion to crickets and adjudged the haemolymph to be the main site of detoxification. (Arnold 1974) was of the opinion that haemocytes do not function in direct defense against the toxicants. (Rakitin 1974) found that haemocytes absorbed a considerable proportion of an administered radioactive insecticide, but he did not say how this affected the insects' response to the insecticide. (Feir 1979) reviewed the cellular and humeral responses of haemocytes to toxic substances. In this paper, the pathological and numerical changes in the haemocyte population of American cockroaches after treatment with malathion are described and the route of insecticide movement inside the insect body is discussed.

MATERIALS AND METHODS

Malathion (technical grade) was made up as a 2% solution in benzene.

Aliquots of 10 μ l of this solution were applied separately to the forefemur, dorsal thoracic terga and ventral abdominal segments, in nonmelanized newly emerged adults (1 hour after eclosion) of both sexes. A metathoracic leg was severed after 15, 30 and 60 minutes of treatment at the COXO-sternal joint. The haemolymph was quickly drawn up to the 0.1 mark in a Thoma white blood cell dilution pipette and the sample was diluted 100 times with 0.2% glacial acetic acid in Ephrussi and Beadle saline (Buck 1953). The first two or three drops of this solution were discarded, small

*Correspondence and reprint requests.

drops were then put on two Knaubers chambers which were immediately covered by a coverslip. Haemocytes were counted in the outer 4 chambers, each containing 16 squares of each chamber. The mean of the two chambers was taken as the haemocyte concentration. At each interval after treatment, four such sets from three different individuals were repeated to determine the standard error. In this way 12 counts were done at each interval. The haemocytes of both sexes were counted independently. The control counts were made in a similar fashion.

The pathological changes in the haemocytes were observed by preparing haemolymph smears after the same time intervals taken for the total counts. The haemocytes were fixed in 5% formalin and stained by the rapid Giemsa method.

The quantitative determination of the haemocyte aberrations viz. mitotic counts was also taken into account. Nuclear and cell diameters of control and treated haemocytes (plasmotocytes) were calculated by ocular micrometer and have been expressed in m/u.

RESULTS AND DISCUSSION

The observations showed that females have significantly more haemocytes than males under the conditions of this experiment (Table 1). The total haemocyte count fell in each time interval after treatment in both sexes (Table 1).

Various cytopathological changes exhibited by haemocytes after malathion treatment included shrinkage of granulocytes, cytoplasmic vacuolization in plasmotocytes (Fig. 2) and mitotic divisions in prohaemocytes (Fig. 3), irregular shapes (Fig. 4) as compared to controls (Fig. 1). A significant reduction in the nuclear diameter and a non-significant reduction in the cell diameter was observed after 60 minutes of treatment (Table 2). The percentage of mitotic figures was increased after 60 minutes of treatment (Table 3).

A fall in total haemocyte count in both the sexes after malathion treatment suggests that the haemocytes might have been used up in the process of detoxification. A substantial fall in numbers was also seen in the controls by 60 minutes. Parathion (Patton 1961) and chlordane (Gupta and Sutherland 1968) had been found to cause a fall in total haemocyte count. (Witten 1969) suggested that the detoxification process

Table 1. Haemocyte counts of *P. americana* after treatment with 2% malathion in benzene.

Place of Application	Time interval	Sex	Control* cells/mm ³		Treated* cells/mm ³
Forefemur	15 min	Male	34	± 1.0	20
		Female	40	± 4.4	21.5 ± 1.5
	30 min	Male	32	± .39	24.7 ± 1.1
		Female	40	± 4.4	21.5 ± 1.5
	60 min	Male	21.6	± 6	14.2 ± 1.2
		Female	28.6	± 1.1	20.9 ± 1.8
Dorsal Thoracic Terga	15 min	Male	50.8	± .96	26.8 ± 1.20
		Female	67.4	± 5.9	42.3 ± 1.1
	30 min	Male	27.7	± 1.8	20.6 ± .08
		Female	60.3	± 1.6	52 ± 14.7
	60 min	Male	28.2	± 1.1	14.8 ± 1.7
		Female	26.8	± .3	21.1 ± 1.7
Central Abdominal Segment	15 min	Male	37.1	± 11.5	28 ± 1.0
		Female	43.6	± 1.5	32.3 ± 3.1
	30 min	Male	25.7	± 1.4	20.5 ± 2.4
		Female	59.9	± 2.0	37.5 ± 1.0
	60 min	Male	28.0	± 3.5	20.3 ± 3.0
		Female	26.4	± 1.4	23.9 ± 1.9

*Round off figures to nearest 1000.

NS = Non-significant.

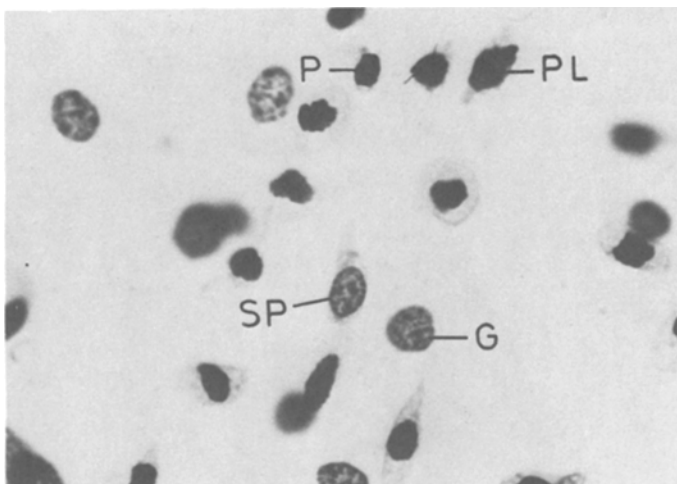


Fig.1 Normal haemocytes showing Prohaemocytes (P) Plasmotocyte (PL) Spherulocyte (SP) Granulocyte (G) x 400.

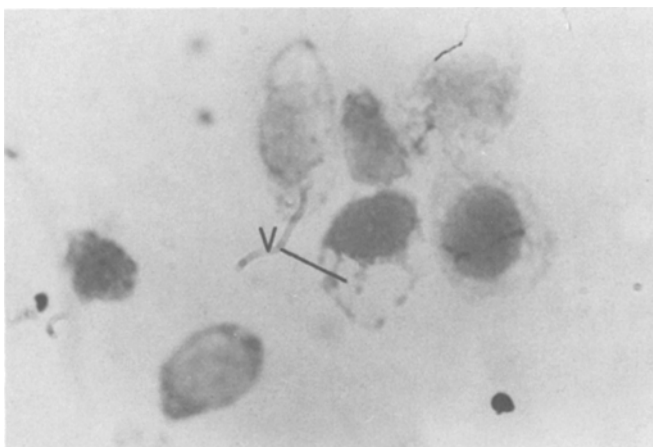


Fig.2 Treated insect showing Vacuolization in Plasmotocytes (V) x 950.

that sets in after insecticidal penetration through the tarsal cuticle caused a fall in total haemocyte population.

Cytopathological changes in haemocytes of Prodenia after arsenical and mercuric chloride poisoning (Yeager and Munson 1942) included rounding up of cells, cell agglutination and increased mitotic frequency. Such changes were also found in the present investigation, and can be interpreted as indicating the passage of malathion to the target organ (Saxena and Saxena 1984) through the haemolymph. Onset of the changes

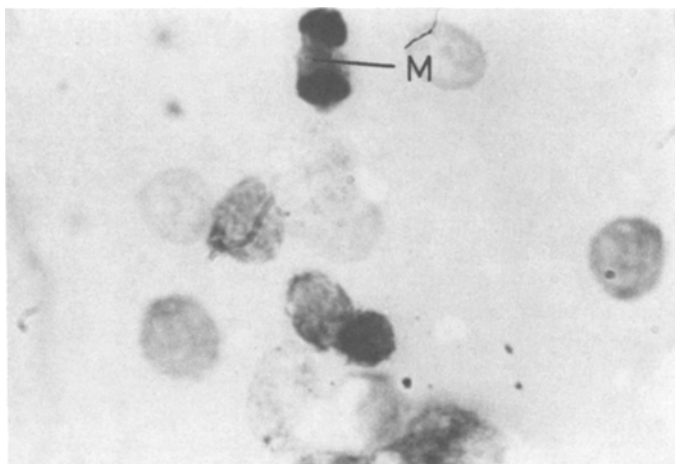


Fig.3 Treated insect showing Mitotic division in Prohaemocytes (M) x 950.

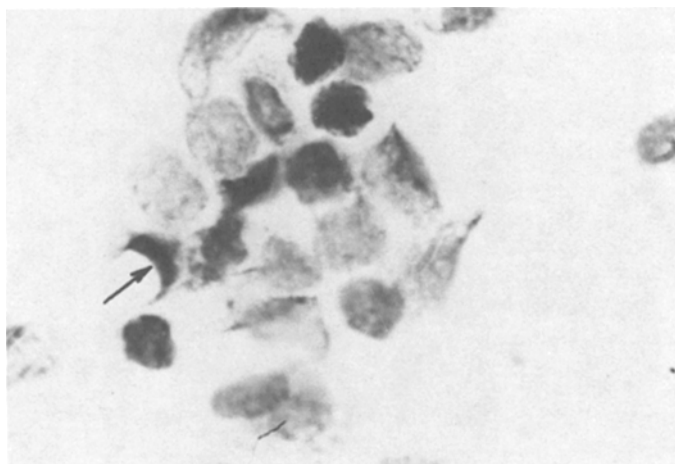


Fig.4 Treated insect showing change in shape of haemocytes (arrow) x 950.

was observed at the time paralytic symptoms appeared, namely 15 minutes after treatment. Based on these findings the present authors do not accept the view of (Gerolt 1969) that penetration of topically applied chemicals is solely via tracheae. Perhaps Gerolt might not have examined the effect of insecticides on haemocytes. Further evidence supporting the passage of malathion through the haemolymph to the target organs was the presence of malathion on thin layer chromatograms and gas liquid chromatograms of the haemolymph after 5 minutes of treatment (Saxena 1982).

Table 2. Quantitative determination of nuclear and cell diameter of plasmatocytes after treatment with 2% malathion (in benzene).

Total No. of plasmatocytes*	Time interval (min)	Nuclear Diameter (m/u)		Cell Diameter (m/u)	
		Control	Treated	Control	Treated
10	60	3.62 \pm .067	3.35 \pm .06*	6.71 \pm .38	6.46 \pm .26**
10	60	3.62 \pm .058	3.34 \pm .06*	6.72 \pm .36	6.44 \pm .22**
10	60	3.63 \pm .070	3.32 \pm .05*	6.73 \pm .34	6.45 \pm .25**

*3 sets were taken in each case.

*(P < .001)

** (P > .05)

Table 3. Mitotic counts of prohaemocytes in control (benzene) and treated (malathion 2%) cockroaches.

Number of Cells counted	Mitotic figures in (Percentage)	
	Control	Treated
50	4%	34%
50	2%	32%
50	2%	38%

Earlier (Burt and Lord 1968; Burt *et al.* 1971) found no evidence for involvement of tracheae in the penetration of pyrethrin I. (Moriarity and French 1971; Polles and Vinson 1972; Olson 1973) had similar findings and demonstrated that lateral spreading of the toxicant in the epicuticular lipids is unimportant and favoured haemolymph to be the chief distributor of toxicant.

AKNOWLEDGEMENTS: Financial assistance to one of us (PNS) by UGC (New Delhi) is gratefully acknowledged.

REFERENCES

- Arnold JW (1974) The haemocytes of insects PP 201-54
In M. Rockstein (ed) The Physiology of Insects
Vol.5, 2nd ed. Academic Press New York.
- Buck JB (1953) Physical properties and chemical
composition of insect blood. Insect Physiol Ed
by Roeder KB, John Wiley New York 147-190.
- Burt PE and Lord KA (1968) The influence of penetra-
tion, distribution, sorption and decomposition on
the poisoning of the cockroach Periplaneta americana
treated topically with diazinon. Entomol Exp Appl
11: 55-67.
- Burt PE, Lord KA, Forrest JM and Goodchild RE (1971)
The spread of topically applied pyrethrin I from
the cuticle to the central nervous system of the
cockroach Periplaneta americana. Entomol Exp Appl
14: 255-269.
- Feir D (1979) Cellular and humoral responses to toxic
substances 415-421 Insect haemocytes Ed. A.P. Gupta
Cambridge University Press, Cambridge, London, New
York, Melbourne.
- Gerolt P (1969) Mode of entry of contact insecticides
J Insect Physiol 15: 569-580.
- Gupta AP and Sutherland DJ (1968) Effects of sublethal
doses of chlordane on the haemocytes and midgut
epithelium of Periplaneta americana. Ann Entomol
Soc Amer 61(4): 910-18.
- Moriarity F and French MC (1971) The uptake of
dieldrin from the cuticular surface of dieldrin
from the cuticular surface of Periplaneta americana
L. Pestic Biochem Physiol 1: 286-292.
- Olson WP (1973) Dieldrin transport in the Insect; An
examination of Gerolt's hypothesis. Pestic Biochem
Physiol 3: 384-392.
- Patton RL (1961) The detoxication function of insect
haemocytes Ann Entomol Soc Amer 54: 696-698.

Polles SG and Vinson SB (1972) Penetration, distribution and metabolism of 14 C- endrin in resistant and susceptible tobacco budworm larvae. J Agr Food Chem 20: 38-41.

Rakitin AA (1974) Adsorption of certain toxicants by various insect tissues. Zh Obstrch Biol 35(1): 127-33.

Saxena Prabhu Narain (1982) Ph.D. thesis entitled "Studies pertaining to effectiveness implying penetration movement and effect on target site of pesticides into insect body with some reference to electronmicroscopy and biochemistry of brain" University of Rajasthan, Jaipur, India.

Saxena SC and Saxena Prabhu N (1984) Ultrastructural study of Periplaneta brain, the possible site of action of malathion. Ind J Exp Biol 22: 169-71.

Witten JM (1969) Haemocyte activity in relation to epidermal cell growth cuticle secretion and cell death in a metamorphosing cyclorrhophan paper. J Insect Physiol 15: 763-78.

Yeager JF and Munson SC (1942) Changes induced in the blood cells of the southern army worm (Prodenia eridania) by the administration of poison. J Agric Res 64(6): 307-32.

Received May 25, 1984; accepted July 11, 1984.